In the Specification:

Please amend the specification as shown:

Please delete the paragraph on page 4, lines 8-13, and replace it with the following paragraph:

Figure 1 shows the cDNA (SEQ ID NO: 3) and translated amino acid (SEQ ID NO: 2) sequences of human brain LysoPLA. The cDNA and amino acid residues that are different between the human and mouse enzymes are in bold, the catalytic triad is marked by *. The primers that were used to clone the human enzyme are indicated by the lines above the cDNA sequences, and the sequences refer to the sequences in the experimental procedures. The 5' and 3' untranslated cDNA sequences were obtained from GenBank.

Please delete the paragraph on page 10, lines 11-19, and replace it with the following paragraph:

Human brain messenger RNA (Clonetech) was used to synthesize first-strand cDNA at 37°C for 1 h using Moloney murine leukemia virus reverse transcripts (Stratagene) and random primers (Promega). Aliquots of the first-strand cDNA were used to amplify the hLysoPLA gene by PCR using Taq DNA polymerase (Promega). The sequences of the primers used in the PCR cycles were: 5 P-GGG GGG CAT ATG TGC GGC AAT AAC ATG TCA ACC CC-3 P (SEQ ID NO: 4) and 5 P-GCG CGA ATT CTC AAT CAA TTG GAG GTA GGA GTT TAT-3 P (SEQ ID NO: 5). These primers were designed based on the human sequences found in the GenBank using the mouse LysoPLA sequences as the probe. Restriction enzyme sites (NdeI and EcoRI) were included near the 5 P-end to facilitate directional cloning (see Fig. 1).

Please delete the paragraph on page 10, line 27, to page 11, line 24, and replace it with the following paragraph:

To clone the human gene, the purified PCR product was ligated into the pCR2.1 vector (Original TA Cloning Kit from Invitrogen) using T4 DNA ligase (Pharmacia) at 14°C for 19 h. The ligase was heat-inactivated by incubating at 70°C for 10 min. The ligation product was used to transform Escherichia coli INVKF P cells, and then plated on LB-Amp (100μg/ml) with X-Gal. After incubation overnight at 37°C and then 6 h at 4°C, several white colonies were selected for overnight cultures in LB-Amp (100μg/ml). The plasmids prepared from the over-night cultures were screened by restriction enzyme analysis using EcoRI and NdeI (Pharmacia). A majority of the screened colonies contained the vector with the correct size insert. The DNA sequence of the insert was obtained using an automated DNA sequencer (Applied Biosystems 373 from Perkin-Elmer). To subclone hLysoPLA from the pCR2.1 vector into the protein